

# Using model systems to understand errant plasticity mechanisms in psychiatric disorders

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***In vivo* model systems are a critical tool for gaining insight into the pathology underlying psychiatric disorders. Although modern functional imaging tools allow study of brain correlates of behavior in clinical groups and genome-wide association studies are beginning to uncover the complex genetic architecture of psychiatric disorders, there is less understanding of pathology at intervening levels of organization. Several psychiatric disorders derive from pathological neural plasticity, and studying the mechanisms that underlie these processes, including reinforcement learning and spike-timing-dependent plasticity, requires the use of animals. It will be particularly important to understand how individual differences in plasticity mechanisms at a cellular level confer resilience on some but lead to disease in others.**

Understanding the neurobiology of psychiatric disorders requires the use of animals. Functional MRI in patient groups has opened a window into the human brain that provides insight into the networks that may be disrupted in various disorders, and positron-emission tomography can show differences in neurotransmitter systems between patient groups and comparison participants<sup>1</sup>. However, this work only suggests gross anatomical correlates of disorders and provides little insight into the neural mechanisms that are disrupted in patient groups. Some psychiatric disorders are likely due to disordered learning caused by pathology in cellular plasticity mechanisms. Insight into disordered plasticity mechanisms at the cellular and synaptic levels requires experiments in model systems.

While it is clear that animals are necessary to gain an understanding of the mechanistic underpinnings of psychiatric disorders, model systems can be used in many ways<sup>2</sup>. One approach to using animals in psychiatric research, which has failed to yield progress, is to develop an animal model of the disease using a behavior thought to be relevant to the disease in conjunction with a manipulation that affects this behavior. The model is then used as a screen in drug discovery by showing that a drug can ameliorate a behavioral deficit in the model<sup>3,4</sup>. In this approach the model's translational utility is assessed with face, construct and predictive validity<sup>5</sup>. Face validity refers to similarity between behavioral or neural features of a disease and the animal model. As there are no well-validated biomarkers of psychiatric disorders<sup>5</sup>, behaviors are usually used to achieve face validity in animal models. However, reducing psychiatric disorders to a single or even a few behaviors is problematic for multiple reasons, including heterogeneity among patients with a single diagnosis.

Construct validity refers to the technique used to generate the animal model. If the neuropathological process that gives rise to the disease was understood, and if that process could be recapitulated in an animal model, this would provide strong construct validity. The advent of transgenic rodent models suggested an approach to generating a model system with good construct validity<sup>6</sup>. Specifically, if a candidate gene existed for a psychiatric disorder, the disease-causing allele could be knocked into a rodent. However, it is now clear that psychiatric disorders result from both a large number of common alleles with low penetrance and a much smaller number of very rare alleles with high penetrance<sup>7,8</sup>. In schizophrenia, each mechanism contributes similarly to disease liability in the population<sup>9</sup> and therefore any single gene accounts for a limited fraction of disease cases. Studying the highly penetrant low-frequency alleles<sup>10</sup> may provide insight into general disease mechanisms. However, expression of a gene that leads to disease in the human brain often recapitulates only a subset of disease features in animal models<sup>11</sup>, therapies that ameliorate deficits in animal models often fail to translate to humans<sup>12</sup> and genes that have clear phenotypes in animal models<sup>3</sup> often have weak penetrance in human populations.

Predictive validity is the ability of the model to predict whether a new treatment will prove effective in the clinical setting. This is often assessed by determining whether current state-of-the-art treatments are effective in the animal model, although effectiveness is often relative to a behavior that itself lacks sufficient validity. Overall, the validity framework appeared plausible but has been ineffective: the clinical efficacy of antipsychotic medications has not improved over the past 50 years of drug development<sup>13,14</sup>.

## Identifying mechanisms in model systems

Animal models can also be used to study the neural mechanisms that underlie behavior or behavioral dimensions<sup>3,15,16</sup>. A detailed understanding of pathology in neural mechanisms relevant to psychiatric disorders can give rise to treatments. Going from an understanding of mechanism to treatment is a difficult step in itself, because tools currently available to treat psychiatric disorders are blunt instruments. For example, even in disorders caused by single

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gene mutations like Huntington's disease<sup>8</sup>, discovery of the gene has led to increased understanding of the underlying pathology but not improvements in treatment. In addition, for this approach to work there needs to be a close collaboration between clinicians and basic scientists, so that highly similar behaviors that are mediated by very similar neural circuits and dynamics within those circuits are studied across species.

Understanding mechanism can, however, lead to improvements in clinical practice. For example, the use of deep-brain stimulation to treat Parkinson's disease was developed from an understanding of how the basal ganglia drives movements<sup>17,18</sup>. Specifically, increased activity in the excitatory subthalamic nucleus (STN) drives increased activity in the inhibitory globus pallidus internal segment (GPi), and this leads to decreased activity in thalamocortical circuits<sup>19</sup>, which drive movements. Although current understanding of this system is more complex<sup>20</sup>, this simple model does predict the effects of manipulations of this circuit on behavior<sup>21</sup>. Lesions of the STN were shown to ameliorate motor deficits in the macaque MPTP model (which results in loss of dopamine neurons) of parkinsonism<sup>22</sup>. Subsequent work in patients examined stimulation of either GPi or the STN and found that both could alleviate symptoms<sup>23</sup>. While deep-brain stimulation is clinically effective, its specific mechanism of action is still not clear<sup>24</sup>. Although this example shows that understanding disease mechanisms can lead to improvements in treatment, the pathophysiology that underlies Parkinson's disease has been understood since the beginning of the 20th century<sup>25</sup>. In psychiatric disorders we still lack understanding of the pathophysiology<sup>26</sup>.

### Neuropsychiatric diseases as pathological learning

Understanding pathogenesis in psychiatric disorders is difficult because it requires establishing causal relationships between events that take place at different levels of scale in the brain. Fully understanding the causal chain of events leading to pathology requires understanding how genetic mutations<sup>7,27</sup> produce changes in development<sup>28–30</sup> that interact with the environment<sup>31</sup> to alter the physiology of neurons and synapses<sup>32</sup>, changing the patterns of activity generated by networks<sup>33–35</sup> and leading to distortions of perception and cognition<sup>36,37</sup>. We know most about disease at the two extremes of this sequence. What is lacking is a mechanistic understanding of events at intervening levels of scale, where pathogenesis proceeds by changing how neurons and synapses work.

Evidence at genetic and behavioral levels suggests that psychiatric disorders may be disorders of learning. For example, many of the genetic loci associated with schizophrenia are related to synaptic plasticity<sup>7</sup>. In addition, at a behavioral level, drug addiction follows from a pathological association of drug cues and motivational systems, and post-traumatic stress disorder (PTSD) develops when the fear and anxiety driven by a traumatic event does not extinguish. Furthermore, across several disorders, treatments currently being put forward from a mechanistic point of view target NMDA receptors, which are closely related to synaptic plasticity. What is missing, however, is an understanding of the pathology in cellular mechanisms that link genetic predispositions, when they exist, to the deficits in learning at a behavioral level. Disorders in dopamine-driven reinforcement learning<sup>38</sup> and spike-timing-dependent plasticity<sup>39</sup> may underlie pathology in, for example, addiction and schizophrenia. Therefore, close links between genetics, plasticity at a cellular level driven by one of these learning mechanisms, and behavior need to be worked out. Below we consider where studying learning and plasticity in model systems has led to suggestions for novel therapeutics and also consider areas where additional research on these mechanisms is

needed. In particular, understanding of how individual differences in plasticity mechanisms confer resiliency on some and lead to disease in others is just beginning in addiction but has been little explored in other disorders.

### Relevance of fear learning and extinction to anxiety disorders

Fear conditioning or, more specifically, threat conditioning<sup>40</sup>, and its extinction is one of the better-understood behaviors with relevance to psychiatric disorders in systems neuroscience. In the fear conditioning model, a neutral conditioned stimulus (CS) is paired with an unconditioned stimulus (US), which often takes the form of a foot shock<sup>41</sup>. Following a few pairings, the animal forms an association between the CS and the US, such that when the animal is brought back in a subsequent session, presentation of the CS leads to freezing. Considerable work has shown that circuits within the amygdala play a critical role in the association of the CS and the US<sup>42–44</sup>. For example, before the association between the CS and the US, cells in the lateral nucleus of the amygdala (LA) respond only weakly to the CS. However, after the association, the CS drives a robust response in LA neurons<sup>45,46</sup>. Furthermore, the increased response to the CS is largely driven by plasticity in the LA, rather than being inherited from other structures<sup>47,48</sup>. The LA projects directly to the central nucleus (CN) of the amygdala, as well as to the basal nucleus, which also projects to the CN<sup>49</sup>. Freezing following presentation of the CS is driven by descending outputs from the CN of the amygdala to the periaqueductal gray.

Fear extinction is the process of extinguishing the behavioral response to the CS. In the extinction paradigm, once a fear association has been formed, it is extinguished by exposing the animal to the CS in the absence of the US. Following a sufficient number of trials, the freezing response to the CS disappears. While extinction learning has often been thought of as unlearning or erasing the original fear association<sup>50</sup>, it is now accepted that most extinction learning (but see<sup>51</sup> and below) is the learning of a new inhibitory association between the CS and no-US, which competes with the original CS–US association<sup>52,53</sup>. This is based on several findings in extinction experiments. Specifically, under renewal, when the CS is extinguished outside of the original context, a return to the original context leads to a return of freezing. Under reinstatement, if the US is delivered by itself following extinction, the CS will subsequently drive freezing. Under recovery, the simple passage of time leads to the reemergence of the fear memory. In addition, after extinction, reassociation of the CS and US in a subsequent conditioning session occurs more quickly, a process referred to as savings. Thus all of these conditions show that the original CS–US association was still present in some form following extinction training.

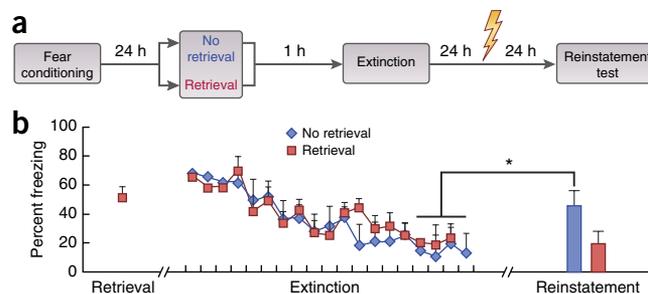
This work has also been translated to human subjects and clinical studies of anxiety disorders<sup>54–58</sup>. Using functional MRI in healthy human participants, it was shown that the neural circuitry underlying fear learning and extinction is shared across humans and rodents<sup>59</sup>. This work has not, however, translated into a useful biomarker for anxiety. In addition, fear conditioning and extinction in animal models differ from the more complex clinical picture of anxiety disorders. For example, participants with diagnosed PTSD that resulted from a single discrete trauma are hyper-responsive to acoustic startle, consistent with the fear conditioning model, whereas patients with PTSD that resulted from cumulative trauma have decreased response to acoustic startle<sup>60</sup>. Discrete-trauma PTSD patients also respond more readily to exposure therapy than cumulative-trauma patients<sup>61</sup>. In addition to this clinical distinction, fear is an emotional state that can only be assessed in human participants, and fear as an emotion

differs from physiological responses to threat assessed in animals<sup>40</sup>. Meta-analyses suggest that patients with anxiety disorders have a modest increase in acquisition of fear associations, particularly a generalization to safety cues (i.e., CSs not associated with aversive USs) as well as a modest increase in expression of fear during extinction<sup>54</sup>. Like functional imaging data, however, behavioral studies of fear learning are not diagnostic of anxiety disorders because there is a large overlap in behavior across groups. In addition, and perhaps most importantly, individual differences in synaptic plasticity at a cellular level that may underlie resilience in some but result in anxiety disorders in others are not known.

Exposure therapy, derived from fear extinction, is an important component of cognitive behavioral therapy in anxiety disorders<sup>57,62</sup>. Similarly to extinction in rodents, extinction in anxiety disorders leads to temporary relief from symptoms, but often fear returns after the passage of time due to renewal or recovery<sup>63</sup>. This has led to a search for ways to improve exposure therapy by enhancing learning. Two ideas for improvement have come from basic science studies of extinction learning<sup>64</sup>. Specifically, the use of D-cycloserine (refs. 65,66) and reconsolidation–extinction to enhance extinction learning<sup>67,68</sup>.

Early experiments showed that blocking NMDA receptors in the amygdala, which interferes with plasticity mechanisms, decreased extinction learning in rodents<sup>69</sup>. It was subsequently shown that D-cycloserine, a noncompetitive NMDA agonist, administered systemically or in the amygdala, could enhance extinction<sup>70</sup>. Follow-up studies showed that D-cycloserine administered to human participants with acrophobia in combination with exposure therapy led to increased extinction learning<sup>71</sup>. Subsequent results have been mixed and current meta-analyses suggest that results are inconclusive, but additional studies should be done to clarify effects and mechanism<sup>72</sup>. Better understanding of the cellular mechanisms that underlie extinction may lead to better treatments to improve extinction learning.

The second approach that has emerged from basic research to increase the effectiveness of exposure therapy is the reconsolidation–extinction paradigm. As discussed above, standard extinction protocols do not lead to erasure of fear memory. Rather they lead to the creation of a new CS–no-US memory that inhibits the original CS–US association. However, when memories are retrieved they become labile and must be reconsolidated<sup>73</sup>, a process that requires protein synthesis. This was first exploited in an extinction paradigm in which, during extinction, rats were given a single CS with either anisomycin, which blocks protein synthesis, or artificial cerebrospinal fluid as a control. Freezing to the single CS did not differ by group. However, on the day after the CS was given with anisomycin, presentation of the CS led to less freezing by the rats that had received anisomycin. Unfortunately, anisomycin is toxic in humans. Therefore, a behavioral procedure was developed in an attempt to achieve the same end. In this study, it was shown that if rats were given a single CS presentation either 10 min or 1 h before extinction training, but not 6 or 24 h before extinction training, the return of fear by recovery, reinstatement or renewal was substantially decreased (Fig. 1)<sup>67</sup>. This was taken as evidence that, following the reconsolidation–extinction model, the fear memory was permanently attenuated. Subsequent studies suggested that this approach also worked in healthy human subjects, in a fear-learning model<sup>68</sup>. Considerable additional work has followed this finding<sup>51,74</sup>. Current meta-analyses suggest that reconsolidation extinction reduces the return of fear in healthy human participants but not in rodents<sup>75</sup>. Efficacy of this procedure has not yet, however, been examined in clinical groups. These results sug-



**Figure 1** Reinstatement of fear following the retrieval–extinction procedure. (a) Conditioning paradigm used to induce and extinguish fear under either a standard extinction protocol or a retrieval–extinction protocol in which a fear memory is retrieved and rendered labile 1 h before extinction. (b) Behavioral performance following extinction and reinstatement. Images adapted from ref. 67, AAAS. Error bars are s.e.m.; \* $P < 0.05$ .

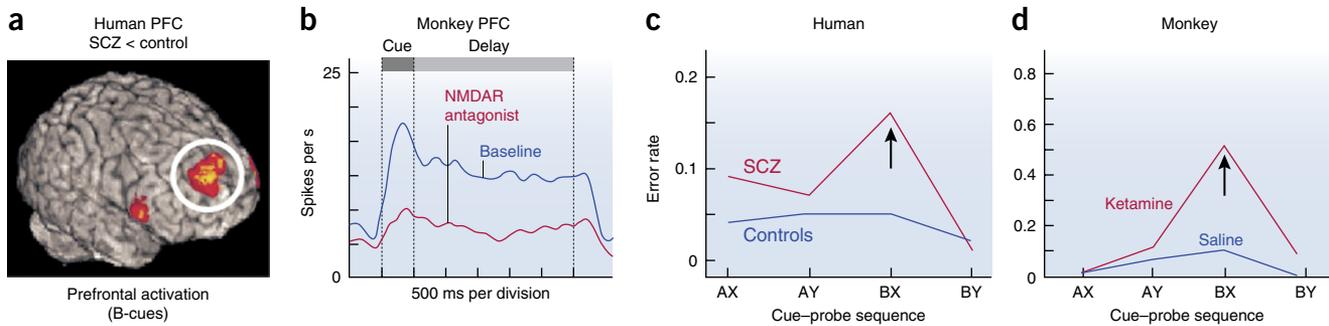
gest that bottom-up work in basic science focused on understanding the neural mechanisms that underlie a relevant behavior can drive treatments based on novel mechanisms.

### Neural circuits underlying compulsive drug taking

The neural circuitry that underlies addiction has considerable overlap with the circuitry that underlies fear learning<sup>76</sup>. Addiction can be considered a pathological association between cues and appetitive USs (i.e., drugs of abuse) just like fear is a pathological association between cues and aversive USs. The amygdala plays an important role in addiction<sup>77</sup>. However, the dopamine system and the striatum are also heavily implicated. Many drugs of abuse either directly or indirectly increase dopamine signaling<sup>78,79</sup>. There is considerable evidence that dopamine codes reward prediction errors (i.e., the difference between the received and expected reward)<sup>80</sup> and that reward prediction errors are an important component of reinforcement learning, or associating stimuli with prediction of reward<sup>81</sup>. This theory predicts that when outcomes are more rewarding than expected, dopamine is released and the estimate of reward associated with cues, actions or the current context are increased. In explicit versions of this model the dopamine signal drives synaptic plasticity on cortical–striatal synapses, and it is these synapses that encode expected reward<sup>82</sup>. In addition, temporal-difference learning models predict that the first cue that predicts reward drives dopamine release<sup>83</sup>. In a simple appetitive Pavlovian experiment, when a CS is first shown, it will not lead to dopamine release because the CS does not predict reward. If a reward (US) is then delivered following the CS, there will be a dopamine response to the reward. However, following learning, when the CS comes to predict the reward, the dopamine response will shift to the CS because it is the first cue that predicts reward. When the reward is subsequently delivered there will be no dopamine response because the reward has been predicted.

Drugs may drive pathological levels of learning because they drive dopamine release and therefore a constant reward prediction error<sup>84</sup>. Under normal physiological conditions, after learning, the reward prediction would equal the received reward (for example, the sight of a food pellet is a cue that predicts the actual reward obtained when the food pellet is consumed) and the reward prediction error would be zero. With no reward prediction error there would be no dopamine release and no additional learning. However, with drugs of abuse that cause an artificial increase in dopamine levels, there would always be a positive reward prediction error and therefore learning. This would cause value estimates to be driven to pathological levels. Although several general aspects of this model have been





**Figure 3** NMDAR antagonists in monkeys replicate both behavioral and neurophysiological features of cognitive deficits in patients with schizophrenia. (a) Blood-oxygen-level dependent (BOLD) signal reflecting maintenance of the B-cue (which countermands the habitual response to subsequent X-probes) in working memory is reduced in prefrontal cortex of schizophrenia patients relative to controls performing the AX-CPT. (b) Focal iontophoretic application of an NMDAR antagonist in prefrontal cortex of monkeys performing the ODR task reduces the spatially selective persistent firing of a prefrontal neuron associated with maintenance of the cue location in spatial working memory. (c) Patients with schizophrenia (red) make selectively more errors on BX-trials relative to controls (blue) on the AX-CPT task. (d) In monkeys performing the AX-CPT, systemic injections of ketamine (an NMDAR antagonist) induce the same error pattern: a selective increase in errors on BX-trials (red line) relative to control (saline injections; blue)<sup>140</sup>. Images in **a** and **c** adapted from ref. 34, American Psychiatric Association; images in **b** adapted from ref. 132, Elsevier; images in **d** adapted from ref. 140, Nature Publishing Group.

### Model systems for understanding cortical network dysfunction in schizophrenia

Schizophrenia is a more complex disorder to study than anxiety or addiction. In anxiety there are better-developed models of the circuitry that gives rise to defensive or avoidance behaviors and in addition there is a reasonable understanding of the pathogenesis. The gap between understanding and disease is larger in schizophrenia. Nonetheless substantial advances have been made in understanding the neurobiology of the disease. Functional imaging<sup>34,35</sup>, magnetoencephalography<sup>102</sup> and electroencephalography<sup>103</sup> have shown changes in brain activity and synchrony patterns<sup>104,105</sup> in patients. These studies provide important insight into pathophysiological signatures of schizophrenia. However, they cannot resolve brain activity in patients at the level of action potentials in neurons. This is a significant limitation because accumulating evidence suggests that one aspect of disease pathogenesis in schizophrenia may involve the molecular mechanisms of activity-dependent synaptic plasticity<sup>106</sup>. Specifically, some risk mutations in schizophrenia involve a cluster of genes that play a role in NMDA receptor (NMDAR)-mediated synaptic transmission<sup>107</sup>. NMDA receptors play a crucial role in triggering activity-dependent synaptic plasticity<sup>108</sup>, and the plasticity mechanism mediated by NMDARs is exquisitely sensitive to the precise timing of action potentials in pre- and postsynaptic neurons<sup>39</sup>. Additional risk mutations involve genes that play a role in the immune system<sup>7</sup>, and new evidence suggests these genes support molecular signaling pathways that target synapses for elimination during development, a process which itself is activity-dependent<sup>109</sup>. This suggests that schizophrenia pathogenesis could involve a disturbance in the linkage between activity patterns at the level of spiking neurons and the strength of synaptic connections in cortical networks. Support for this class of theories could significantly redirect strategies to identify novel treatments. However, because pathogenic mechanisms of this type depend crucially on the timing of action potentials in synaptically coupled neurons and the disruption of this timing in the disease state, testing them requires resolving electrical activity at the cellular level, which requires the use of animals.

To effectively model the interaction between activity patterns and synaptic plasticity mechanisms, it is necessary to first try to replicate pathophysiological activity patterns likely to occur in the human disease in the model system. This will be more likely if behavior is well matched between patients with schizophrenia and the model system.

Consequently, the two species should perform the same behavioral task. In addition, a manipulation should be identified that causes the emergence of the same behavioral error pattern in the animal model that is evident in patients. Second, because neural activity patterns are constrained by patterns of anatomical connectivity in cortical networks, it is optimal if the neural system mediating performance in the animal is as similar in organization as possible to the corresponding network in the human brain. As behavioral and neurophysiological deficits in schizophrenia often preferentially involve the prefrontal cortex<sup>110,111</sup>, this places particular weight on the necessity to optimize homology of prefrontal networks across species, and this criterion favors primate over rodent models<sup>112,113</sup>.

This raises the question of which behavioral features of schizophrenia provide the most promising targets for successful translation to animals. The clinical symptoms of schizophrenia, which include hallucinations, delusions and flatness of affect<sup>114</sup> present obvious challenges for translation to animals (not least of which is that these symptoms are typically evaluated clinically by talking to patients). However, patients with schizophrenia also exhibit deficits in cognitive functions that are measurable using automated behavioral tasks that can be translated to animal models. Understanding the neural basis of cognitive deficits rather than clinical symptoms of schizophrenia has therefore been the focus of translational research<sup>31,115</sup>. Several multi-investigator consortia have identified behavioral models that most reliably measure specific cognitive deficits reflecting genetic risk in schizophrenia, including deficits in executive control, working memory, attention and sensory gating<sup>116–119</sup>. As with the other psychiatric disorders, reducing a disease as complex as schizophrenia to one or a few cognitive deficits in an animal model will likely lead to a model with limited validity, and therefore results from these studies should be interpreted carefully.

Patients with schizophrenia, and to a lesser degree their first-degree relatives, exhibit deficits in tasks that require spatial working memory<sup>120</sup>. A deficit in spatial working memory can be demonstrated in patients performing the oculomotor delayed response (ODR) task<sup>121</sup>. In this task, subjects direct a saccadic eye movement to the location of a peripheral visual cue several seconds after it disappears. Working memory for the location of the cue is associated with the persistent activation of spatially selective neurons in the dorsolateral prefrontal cortex in monkeys<sup>122,123</sup>. Dorsolateral prefrontal lesions<sup>124</sup>, inactivation<sup>125</sup> or injection of D1 dopamine receptor antagonists

into monkey prefrontal cortex<sup>126</sup> induce spatially restricted deficits in working memory. Importantly, performance on the ODR task is preserved after insult to prefrontal cortex if the target remains visible through the delay, providing a visual target for the saccade at the time it is executed. This establishes the specificity of the deficit for working memory. The ODR task was successfully back-translated from monkeys to patients with schizophrenia and first-degree relatives to demonstrate a specific impairment in spatial working memory<sup>127</sup>. It has been repeatedly documented, using functional imaging in patients, that reduced performance on the ODR and other working memory tasks is associated with reduced activation of prefrontal cortex<sup>128,129</sup> (Fig. 3a).

Most efforts to model behavioral and neurophysiological features of schizophrenia in nonhuman primates have been based on administration of noncompetitive NMDAR antagonists such as ketamine or phencyclidine<sup>130</sup>. Blocking NMDAR in human control subjects replicates positive and negative symptoms as well as some cognitive deficits of schizophrenia<sup>131</sup>. In monkeys, local iontophoretic application of NMDAR antagonists disrupts the persistent activation of prefrontal neurons associated with spatial working memory<sup>132</sup> (Fig. 3b). Other studies have shown that systemic administration of ketamine to monkeys induces errors in a rule-based antisaccade task similar to those produced by patients, and neural recording has characterized concomitant reduction in the encoding of task-related information by prefrontal neurons<sup>133–135</sup>. Collectively, these studies provide important insight into how prefrontal circuits computationally fail in response to loss of NMDAR function. However, some robust findings in patients are not replicated by the NMDAR model<sup>136</sup>.

The AX-CPT (continuous performance task) is a task that measures specific deficits in executive control in patients with schizophrenia<sup>34,137–139</sup> and has been successfully translated to monkeys<sup>140,141</sup>. In this task, a cue stimulus modifies the response required to a subsequent probe. On some trials (BX trials), the cue stored in working memory countermands a habitual or prepotent response to the probe, and it is specifically on this subset of trials that patients with schizophrenia commit the most errors (Fig. 3c)<sup>117,137,138</sup>. Administration of ketamine to monkeys performing the AX-CPT induces the same error pattern (Fig. 3d)<sup>140</sup>. This is one example of a strong behavioral match between the error pattern of patients and an animal model.

However, the fact that a drug in monkeys is able to mimic the symptoms of a disease in humans is not evidence that the drug and disease work through the same neural mechanism. Testing causal theories at the cell and circuit levels requires neural recordings to evaluate the impact of mutations on neural function. The *Df(16)A<sup>+/-</sup>* mouse<sup>142</sup> is engineered to mimic the 22q11.2 chromosomal microdeletion (DiGeorge syndrome), which confers a 30-fold increase in risk of developing schizophrenia and is among the strongest genetic risks for the disease<sup>142</sup>. This mouse exhibits behavioral deficits similar to those seen in schizophrenia, including reduced sensory gating<sup>143</sup> and impaired spatial working memory<sup>143,144</sup>, as well as reduced gamma band synchrony between prefrontal cortex and hippocampus<sup>144</sup>. Deletion of a single gene (*Dgcr8*) in this region that is involved in microRNA processing confers much of the phenotype and has been shown to produce deficits in short-term synaptic plasticity in slice recordings<sup>145</sup>. MicroRNA are involved in the post-transcriptional regulation of other genes. Genome-wide association studies indicate that the large majority of single nucleotide polymorphisms increasing risk of schizophrenia, as with other psychiatric disorders, do not occur in coding regions of genes<sup>8</sup>, suggesting protein structure may be largely normal in the disease. Both observations point to the possibility that a

defect in gene regulation could trigger pathogenesis in schizophrenia, potentially via activity-dependent gene regulatory pathways.

## CONCLUSIONS

It has proven difficult to use animal models to screen for new drugs with therapeutic potential. While models have been generated that satisfy face, construct and predictive validity, they have not facilitated drug discovery. Animals are, however, the only available tool for mechanistic studies of the neural circuitry that drives behavior. More importantly, animals provide the only tool available to study learning driven neural plasticity at a cellular level, and pathology in these plastic processes likely underlies many psychiatric disorders. Understanding individual differences in plasticity mechanisms may further lead to insights into why, given similar genetic backgrounds or behavioral experiences, some individuals develop disorders while others are resilient. This understanding may then lead to better treatments.

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## AUTHOR CONTRIBUTIONS

B.B.A. and M.V.C. wrote the paper.

## COMPETING FINANCIAL INTERESTS

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- Slifstein, M. *et al.* Deficits in prefrontal cortical and extrastriatal dopamine release in schizophrenia: a positron emission tomographic functional magnetic resonance imaging study. *JAMA Psychiatry* **72**, 316–324 (2015).
- Insel, T.R. From animal models to model animals. *Biol. Psychiatry* **62**, 1337–1339 (2007).
- Wong, A.H. & Josselyn, S.A. Caution when diagnosing your mouse with schizophrenia: the use and misuse of model animals for understanding psychiatric disorders. *Biol. Psychiatry* **79**, 32–38 (2016).
- Cosgrove, V.E., Kelseo, J.R. & Suppes, T. Toward a valid animal model of bipolar disorder: how the research domain criteria help bridge the clinical-basic science divide. *Biol. Psychiatry* **79**, 62–70 (2016).
- Nestler, E.J. & Hyman, S.E. Animal models of neuropsychiatric disorders. *Nat. Neurosci.* **13**, 1161–1169 (2010).
- Campbell, I.L. & Gold, L.H. Transgenic modeling of neuropsychiatric disorders. *Mol. Psychiatry* **1**, 105–120 (1996).
- Schizophrenia Working Group of the Psychiatric Genomics Consortium. Biological insights from 108 schizophrenia-associated genetic loci. *Nature* **511**, 421–427 (2014).
- Insel, T.R. & Collins, F.S. Psychiatry in the genomics era. *Am. J. Psychiatry* **160**, 616–620 (2003).
- Gratten, J., Wray, N.R., Keller, M.C. & Visscher, P.M. Large-scale genomics unveils the genetic architecture of psychiatric disorders. *Nat. Neurosci.* **17**, 782–790 (2014).
- Sullivan, P.F., Daly, M.J. & O'Donovan, M. Genetic architectures of psychiatric disorders: the emerging picture and its implications. *Nat. Rev. Genet.* **13**, 537–551 (2012).
- Puzzo, D., Gulisano, W., Palmeri, A. & Arancio, O. Rodent models for Alzheimer's disease drug discovery. *Expert Opin. Drug Discov.* **10**, 703–711 (2015).
- Franco, R. & Cedazo-Minguez, A. Successful therapies for Alzheimer's disease: why so many in animal models and none in humans? *Front. Pharmacol.* **5**, 146 (2014).
- Insel, T.R. Rethinking schizophrenia. *Nature* **468**, 187–193 (2010).
- Lieberman, J.A. & Stroup, T.S. The NIMH-CATIE schizophrenia study: what did we learn? *Am. J. Psychiatry* **168**, 770–775 (2011).
- Fernando, A.B. & Robbins, T.W. Animal models of neuropsychiatric disorders. *Annu. Rev. Clin. Psychol.* **7**, 39–61 (2011).
- Pine, D.S. & Leibenluft, E. Biomarkers with a mechanistic focus. *JAMA Psychiatry* **72**, 633–634 (2015).
- Albin, R.L., Young, A.B. & Penney, J.B. The functional anatomy of basal ganglia disorders. *Trends Neurosci.* **12**, 366–375 (1989).
- DeLong, M.R. Primate models of movement disorders of basal ganglia origin. *Trends Neurosci.* **13**, 281–285 (1990).
- Alexander, G.E., DeLong, M.R. & Strick, P.L. Parallel organization of functionally segregated circuits linking basal ganglia and cortex. *Annu. Rev. Neurosci.* **9**, 357–381 (1986).

20. Cui, G. *et al.* Concurrent activation of striatal direct and indirect pathways during action initiation. *Nature* **494**, 238–242 (2013).
21. Kravitz, A.V. *et al.* Regulation of parkinsonian motor behaviours by optogenetic control of basal ganglia circuitry. *Nature* **466**, 622–626 (2010).
22. Bergman, H., Wichmann, T. & DeLong, M.R. Reversal of experimental parkinsonism by lesions of the subthalamic nucleus. *Science* **249**, 1436–1438 (1990).
23. Obeso, J.A. *et al.* Surgical treatment of Parkinson's disease. *Baillieres Clin. Neurol.* **6**, 125–145 (1997).
24. Wichmann, T. & DeLong, M.R. Deep brain stimulation for movement disorders of basal ganglia origin: restoring function or functionality? *Neurotherapeutics* **13**, 264–283 (2016).
25. Lees, A.J. Unresolved issues relating to the shaking palsy on the celebration of James Parkinson's 250th birthday. *Mov. Disord.* **22** (Suppl. 17), S327–S334 (2007).
26. Kapur, S., Phillips, A.G. & Insel, T.R. Why has it taken so long for biological psychiatry to develop clinical tests and what to do about it? *Mol. Psychiatry* **17**, 1174–1179 (2012).
27. Kirov, G. *et al.* De novo CNV analysis implicates specific abnormalities of postsynaptic signalling complexes in the pathogenesis of schizophrenia. *Mol. Psychiatry* **17**, 142–153 (2012).
28. Marengo, S. & Weinberger, D.R. The neurodevelopmental hypothesis of schizophrenia: following a trail of evidence from cradle to grave. *Dev. Psychopathol.* **12**, 501–527 (2000).
29. Owen, M.J., O'Donovan, M.C., Thapar, A. & Craddock, N. Neurodevelopmental hypothesis of schizophrenia. *Br. J. Psychiatry* **198**, 173–175 (2011).
30. MacDonald, A.W. III & Chafee, M.V. Translational and developmental perspective on N-methyl-D-aspartate synaptic deficits in schizophrenia. *Dev. Psychopathol.* **18**, 853–876 (2006).
31. Burrows, E.L. & Hannan, A.J. Cognitive endophenotypes, gene-environment interactions and experience-dependent plasticity in animal models of schizophrenia. *Biol. Psychol.* **116**, 82–89 (2016).
32. Crabtree, G.W. & Gogos, J.A. Synaptic plasticity, neural circuits, and the emerging role of altered short-term information processing in schizophrenia. *Front. Synaptic Neurosci.* **6**, 28 (2014).
33. MacDonald, A.W. III & Carter, C.S. Event-related fMRI study of context processing in dorsolateral prefrontal cortex of patients with schizophrenia. *J. Abnorm. Psychol.* **112**, 689–697 (2003).
34. MacDonald, A.W. III *et al.* Specificity of prefrontal dysfunction and context processing deficits to schizophrenia in never-medicated patients with first-episode psychosis. *Am. J. Psychiatry* **162**, 475–484 (2005).
35. Yoon, J.H. *et al.* Association of dorsolateral prefrontal cortex dysfunction with disrupted coordinated brain activity in schizophrenia: relationship with impaired cognition, behavioral disorganization, and global function. *Am. J. Psychiatry* **165**, 1006–1014 (2008).
36. Millan, M.J. *et al.* Cognitive dysfunction in psychiatric disorders: characteristics, causes and the quest for improved therapy. *Nat. Rev. Drug Discov.* **11**, 141–168 (2012).
37. Keefe, R.S. & Harvey, P.D. Cognitive impairment in schizophrenia. *Handb. Exp. Pharmacol.* **213**, 11–37 (2012).
38. Dayan, P. & Niv, Y. Reinforcement learning: the good, the bad and the ugly. *Curr. Opin. Neurobiol.* **18**, 185–196 (2008).
39. Feldman, D.E. The spike-timing dependence of plasticity. *Neuron* **75**, 556–571 (2012).
40. LeDoux, J.E. Coming to terms with fear. *Proc. Natl. Acad. Sci. USA* **111**, 2871–2878 (2014).
41. Romanski, L.M. & LeDoux, J.E. Equipotentiality of thalamo-amygdala and thalamo-cortico-amygdala circuits in auditory fear conditioning. *J. Neurosci.* **12**, 4501–4509 (1992).
42. LeDoux, J.E. Emotion circuits in the brain. *Annu. Rev. Neurosci.* **23**, 155–184 (2000).
43. Herry, C. & Johansen, J.P. Encoding of fear learning and memory in distributed neuronal circuits. *Nat. Neurosci.* **17**, 1644–1654 (2014).
44. Davis, M. The role of the amygdala in conditioned and unconditioned fear and anxiety. in *The Amygdala* (ed. Aggleton, J.P.) 213–288 (Oxford University Press, 2000).
45. Quirk, G.J., Armony, J.L. & LeDoux, J.E. Fear conditioning enhances different temporal components of tone-evoked spike trains in auditory cortex and lateral amygdala. *Neuron* **19**, 613–624 (1997).
46. Quirk, G.J., Reppas, C. & LeDoux, J.E. Fear conditioning enhances short-latency auditory responses of lateral amygdala neurons: parallel recordings in the freely behaving rat. *Neuron* **15**, 1029–1039 (1995).
47. Johansen, J.P. *et al.* Optical activation of lateral amygdala pyramidal cells instructs associative fear learning. *Proc. Natl. Acad. Sci. USA* **107**, 12692–12697 (2010).
48. Nabavi, S. *et al.* Engineering a memory with LTD and LTP. *Nature* **511**, 348–352 (2014).
49. Duvarci, S. & Pare, D. Amygdala microcircuits controlling learned fear. *Neuron* **82**, 966–980 (2014).
50. Rescorla, R.A. & Wagner, A.R. A theory of Pavlovian conditioning: variations in the effectiveness of reinforcement and nonreinforcement. in *Classical Conditioning II: Current Research and Theory* (eds. Black, A.H. & Prokasy, W.F.) 64–99 (Appleton-Century-Crofts, New York, 1972).
51. Clem, R.L. & Schiller, D. New learning and unlearning: strangers or accomplices in threat memory attenuation? *Trends Neurosci.* **39**, 340–351 (2016).
52. Bouton, M.E. Context, ambiguity, and unlearning: sources of relapse after behavioral extinction. *Biol. Psychiatry* **52**, 976–986 (2002).
53. Bouton, M.E. Context, time, and memory retrieval in the interference paradigms of Pavlovian learning. *Psychol. Bull.* **114**, 80–99 (1993).
54. Duits, P. *et al.* Updated meta-analysis of classical fear conditioning in the anxiety disorders. *Depress. Anxiety* **32**, 239–253 (2015).
55. Mineka, S. & Oehlberg, K. The relevance of recent developments in classical conditioning to understanding the etiology and maintenance of anxiety disorders. *Acta Psychol. (Amst.)* **127**, 567–580 (2008).
56. Kindt, M. A behavioural neuroscience perspective on the aetiology and treatment of anxiety disorders. *Behav. Res. Ther.* **62**, 24–36 (2014).
57. Graham, B.M. & Milad, M.R. The study of fear extinction: implications for anxiety disorders. *Am. J. Psychiatry* **168**, 1255–1265 (2011).
58. Britton, J.C., Lissek, S., Grillon, C., Norcross, M.A. & Pine, D.S. Development of anxiety: the role of threat appraisal and fear learning. *Depress. Anxiety* **28**, 5–17 (2011).
59. Delgado, M.R., Nearing, K.I., Ledoux, J.E. & Phelps, E.A. Neural circuitry underlying the regulation of conditioned fear and its relation to extinction. *Neuron* **59**, 829–838 (2008).
60. McTeague, L.M. & Lang, P.J. The anxiety spectrum and the reflex physiology of defense: from circumscribed fear to broad distress. *Depress. Anxiety* **29**, 264–281 (2012).
61. Lang, P.J. & McTeague, L.M. Discrete and recurrent traumatization in PTSD: fear vs. anxious misery. *J. Clin. Psychol. Med. Settings* **18**, 207–209 (2011).
62. Pine, D.S. & Klein, R.G. Anxiety disorders. in *Rutter's Child and Adolescent Psychiatry* (eds. Thapar, A. *et al.*) 822–840 (John Wiley & Sons, New York, 2015).
63. Arch, J.J. & Craske, M.G. First-line treatment: a critical appraisal of cognitive behavioral therapy developments and alternatives. *Psychiatr. Clin. North Am.* **32**, 525–547 (2009).
64. Quirk, G.J. *et al.* Erasing fear memories with extinction training. *J. Neurosci.* **30**, 14993–14997 (2010).
65. Rodrigues, H. *et al.* Does D-cycloserine enhance exposure therapy for anxiety disorders in humans? A meta-analysis. *PLoS One* **9**, e93519 (2014).
66. Bowers, M.E. & Ressler, K.J. An overview of translationally informed treatments for posttraumatic stress disorder: animal models of Pavlovian fear conditioning to human clinical trials. *Biol. Psychiatry* **78**, E15–E27 (2015).
67. Monfils, M.H., Cowansage, K.K., Klann, E. & LeDoux, J.E. Extinction-reconsolidation boundaries: key to persistent attenuation of fear memories. *Science* **324**, 951–955 (2009).
68. Schiller, D. *et al.* Preventing the return of fear in humans using reconsolidation update mechanisms. *Nature* **463**, 49–53 (2010).
69. Falls, W.A., Miserendino, M.J. & Davis, M. Extinction of fear-potentiated startle: blockade by infusion of an NMDA antagonist into the amygdala. *J. Neurosci.* **12**, 854–863 (1992).
70. Walker, D.L., Ressler, K.J., Lu, K.T. & Davis, M. Facilitation of conditioned fear extinction by systemic administration or intra-amygdala infusions of D-cycloserine as assessed with fear-potentiated startle in rats. *J. Neurosci.* **22**, 2343–2351 (2002).
71. Ressler, K.J. *et al.* Cognitive enhancers as adjuncts to psychotherapy: use of D-cycloserine in phobic individuals to facilitate extinction of fear. *Arch. Gen. Psychiatry* **61**, 1136–1144 (2004).
72. Ori, R. *et al.* Augmentation of cognitive and behavioural therapies (CBT) with d-cycloserine for anxiety and related disorders. *Cochrane Database Syst. Rev.* **5**, CD007803 (2015).
73. Sara, S.J. Retrieval and reconsolidation: toward a neurobiology of remembering. *Learn. Mem.* **7**, 73–84 (2000).
74. Auber, A., Tedesco, V., Jones, C.E., Monfils, M.H. & Chiamulera, C. Post-retrieval extinction as reconsolidation interference: methodological issues or boundary conditions? *Psychopharmacology (Berl.)* **226**, 631–647 (2013).
75. Kredlow, M.A., Unger, L.D. & Otto, M.W. Harnessing reconsolidation to weaken fear and appetitive memories: A meta-analysis of post-retrieval extinction effects. *Psychol. Bull.* **142**, 314–336 (2016).
76. Peters, J., Kalivas, P.W. & Quirk, G.J. Extinction circuits for fear and addiction overlap in prefrontal cortex. *Learn. Mem.* **16**, 279–288 (2009).
77. Belin, D., Jonkman, S., Dickinson, A., Robbins, T.W. & Everitt, B.J. Parallel and interactive learning processes within the basal ganglia: relevance for the understanding of addiction. *Behav. Brain Res.* **199**, 89–102 (2009).
78. Di Chiara, G. Nucleus accumbens shell and core dopamine: differential role in behavior and addiction. *Behav. Brain Res.* **137**, 75–114 (2002).
79. Wise, R.A. Dopamine and reward: the anhedonia hypothesis 30 years on. *Neurotox. Res.* **14**, 169–183 (2008).
80. Schultz, W. Getting formal with dopamine and reward. *Neuron* **36**, 241–263 (2002).
81. Schultz, W., Dayan, P. & Montague, P.R. A neural substrate of prediction and reward. *Science* **275**, 1593–1599 (1997).
82. Frank, M.J. Dynamic dopamine modulation in the basal ganglia: a neurocomputational account of cognitive deficits in medicated and nonmedicated Parkinsonism. *J. Cogn. Neurosci.* **17**, 51–72 (2005).
83. Hollerman, J.R. & Schultz, W. Dopamine neurons report an error in the temporal prediction of reward during learning. *Nat. Neurosci.* **1**, 304–309 (1998).
84. Redish, A.D. Addiction as a computational process gone awry. *Science* **306**, 1944–1947 (2004).

85. Marks, K.R., Kearns, D.N., Christensen, C.J., Silberberg, A. & Weiss, S.J. Learning that a cocaine reward is smaller than expected: a test of Redish's computational model of addiction. *Behav. Brain Res.* **212**, 204–207 (2010).
86. Volkow, N.D., Fowler, J.S., Wang, G.J., Baler, R. & Telang, F. Imaging dopamine's role in drug abuse and addiction. *Neuropharmacology* **56** (Suppl. 1), 3–8 (2009).
87. Volkow, N.D., Wang, G.J., Fowler, J.S., Tomasi, D. & Baler, R. Food and drug reward: overlapping circuits in human obesity and addiction. *Curr. Top. Behav. Neurosci.* **11**, 1–24 (2012).
88. Belin, D., Belin-Rauscent, A., Murray, J.E. & Everitt, B.J. Addiction: failure of control over maladaptive incentive habits. *Curr. Opin. Neurobiol.* **23**, 564–572 (2013).
89. Janak, P.H. & Tye, K.M. From circuits to behaviour in the amygdala. *Nature* **517**, 284–292 (2015).
90. Namburi, P. *et al.* A circuit mechanism for differentiating positive and negative associations. *Nature* **520**, 675–678 (2015).
91. Stuber, G.D. *et al.* Excitatory transmission from the amygdala to nucleus accumbens facilitates reward seeking. *Nature* **475**, 377–380 (2011).
92. Britt, J.P. *et al.* Synaptic and behavioral profile of multiple glutamatergic inputs to the nucleus accumbens. *Neuron* **76**, 790–803 (2012).
93. Tye, K.M., Stuber, G.D., de Ridder, B., Bonci, A. & Janak, P.H. Rapid strengthening of thalamo-amygdala synapses mediates cue-reward learning. *Nature* **453**, 1253–1257 (2008).
94. White, N.M. Addictive drugs as reinforcers: multiple partial actions on memory systems. *Addiction* **91**, 921–949 discussion 951–965 (1996).
95. Belin-Rauscent, A., Fouyssac, M., Bonci, A. & Belin, D. How preclinical models evolved to resemble the diagnostic criteria of drug addiction. *Biol. Psychiatry* **79**, 39–46 (2016).
96. Olmstead, M.C., Parkinson, J.A., Miles, F.J., Everitt, B.J. & Dickinson, A. Cocaine-seeking by rats: regulation, reinforcement and activation. *Psychopharmacology (Berl.)* **152**, 123–131 (2000).
97. Belin, D., Mar, A.C., Dalley, J.W., Robbins, T.W. & Everitt, B.J. High impulsivity predicts the switch to compulsive cocaine-taking. *Science* **320**, 1352–1355 (2008).
98. Waselus, M. *et al.* Long-term effects of cocaine experience on neuroplasticity in the nucleus accumbens core of addiction-prone rats. *Neuroscience* **248**, 571–584 (2013).
99. Flagel, S.B. *et al.* Genetic background and epigenetic modifications in the core of the nucleus accumbens predict addiction-like behavior in a rat model. *Proc. Natl. Acad. Sci. USA* **113**, E2861–E2870 (2016).
100. Xue, Y.X. *et al.* A memory retrieval-extinction procedure to prevent drug craving and relapse. *Science* **336**, 241–245 (2012).
101. Das, R.K., Freeman, T.P. & Kamboj, S.K. The effects of N-methyl D-aspartate and B-adrenergic receptor antagonists on the reconsolidation of reward memory: a meta-analysis. *Neurosci. Biobehav. Rev.* **37**, 240–255 (2013).
102. Georgopoulos, A.P. *et al.* Synchronous neural interactions assessed by magnetoencephalography: a functional biomarker for brain disorders. *J. Neural Eng.* **4**, 349–355 (2007).
103. Minzenberg, M.J. *et al.* Gamma oscillatory power is impaired during cognitive control independent of medication status in first-episode schizophrenia. *Neuropsychopharmacology* **35**, 2590–2599 (2010).
104. Spellman, T.J. & Gordon, J.A. Synchrony in schizophrenia: a window into circuit-level pathophysiology. *Curr. Opin. Neurobiol.* **30**, 17–23 (2015).
105. Uhlhaas, P.J. & Singer, W. Oscillations and neuronal dynamics in schizophrenia: the search for basic symptoms and translational opportunities. *Biol. Psychiatry* **77**, 1001–1009 (2015).
106. Hall, J., Trent, S., Thomas, K.L., O'Donovan, M.C. & Owen, M.J. Genetic risk for schizophrenia: convergence on synaptic pathways involved in plasticity. *Biol. Psychiatry* **77**, 52–58 (2015).
107. Fromer, M. *et al.* De novo mutations in schizophrenia implicate synaptic networks. *Nature* **506**, 179–184 (2014).
108. Malenka, R.C. & Nicoll, R.A. NMDA-receptor-dependent synaptic plasticity: multiple forms and mechanisms. *Trends Neurosci.* **16**, 521–527 (1993).
109. Sekar, A. *et al.* Schizophrenia risk from complex variation of complement component 4. *Nature* **530**, 177–183 (2016).
110. Goldman-Rakic, P.S. Working memory dysfunction in schizophrenia. *J. Neuropsychiatry Clin. Neurosci.* **6**, 348–357 (1994).
111. Selemon, L.D., Kleinman, J.E., Herman, M.M. & Goldman-Rakic, P.S. Smaller frontal gray matter volume in postmortem schizophrenic brains. *Am. J. Psychiatry* **159**, 1983–1991 (2002).
112. Ahn, S. & Phillips, A.G. Daily monitoring of dopamine efflux reveals a short-lasting occlusion of the dopamine agonist properties of d-amphetamine by dopamine transporter blockers GBR 12909 and methylphenidate. *ACS Chem. Neurosci.* **4**, 817–824 (2013).
113. Preuss, T.M. Do rats have prefrontal cortex? The Rose-Woolsey-Akert program reconsidered. *J. Cogn. Neurosci.* **7**, 1–24 (1995).
114. Goghari, V.M., Sponheim, S.R. & MacDonald, A.W. III. The functional neuroanatomy of symptom dimensions in schizophrenia: a qualitative and quantitative review of a persistent question. *Neurosci. Biobehav. Rev.* **34**, 468–486 (2010).
115. Simen, A.A., DiLeone, R. & Arnsten, A.F. Primate models of schizophrenia: future possibilities. *Prog. Brain Res.* **179**, 117–125 (2009).
116. Nuechterlein, K.H. *et al.* The MATRICS Consensus Cognitive Battery, part 1: test selection, reliability, and validity. *Am. J. Psychiatry* **165**, 203–213 (2008).
117. Carter, C.S., Minzenberg, M., West, R. & MacDonald, A. III. CNTRICS imaging biomarker selections: Executive control paradigms. *Schizophr. Bull.* **38**, 34–42 (2012).
118. Barch, D.M., Moore, H., Nee, D.E., Manoach, D.S. & Luck, S.J. CNTRICS imaging biomarkers selection: Working memory. *Schizophr. Bull.* **38**, 43–52 (2012).
119. Butler, P.D. *et al.* Perceptual measurement in schizophrenia: promising electrophysiology and neuroimaging paradigms from CNTRICS. *Schizophr. Bull.* **38**, 81–91 (2012).
120. Lee, J. & Park, S. Working memory impairments in schizophrenia: a meta-analysis. *J. Abnorm. Psychol.* **114**, 599–611 (2005).
121. Goldman-Rakic, P.S. Cellular basis of working memory. *Neuron* **14**, 477–485 (1995).
122. Funahashi, S., Bruce, C.J. & Goldman-Rakic, P.S. Mnemonic coding of visual space in the monkey's dorsolateral prefrontal cortex. *J. Neurophysiol.* **61**, 331–349 (1989).
123. Chafee, M.V. & Goldman-Rakic, P.S. Matching patterns of activity in primate prefrontal area 8a and parietal area 7ip neurons during a spatial working memory task. *J. Neurophysiol.* **79**, 2919–2940 (1998).
124. Funahashi, S., Bruce, C.J. & Goldman-Rakic, P.S. Dorsolateral prefrontal lesions and oculomotor delayed-response performance: evidence for mnemonic “scotomas”. *J. Neurosci.* **13**, 1479–1497 (1993).
125. Chafee, M.V. & Goldman-Rakic, P.S. Inactivation of parietal and prefrontal cortex reveals interdependence of neural activity during memory-guided saccades. *J. Neurophysiol.* **83**, 1550–1566 (2000).
126. Sawaguchi, T. & Goldman-Rakic, P.S. The role of D1-dopamine receptor in working memory: local injections of dopamine antagonists into the prefrontal cortex of rhesus monkeys performing an oculomotor delayed-response task. *J. Neurophysiol.* **71**, 515–528 (1994).
127. Park, S., Holzman, P.S. & Goldman-Rakic, P.S. Spatial working memory deficits in the relatives of schizophrenic patients. *Arch. Gen. Psychiatry* **52**, 821–828 (1995).
128. Driesen, N.R. *et al.* Impairment of working memory maintenance and response in schizophrenia: functional magnetic resonance imaging evidence. *Biol. Psychiatry* **64**, 1026–1034 (2008).
129. Eryilmaz, H. *et al.* Disrupted working memory circuitry in schizophrenia: disentangling fMRI markers of core pathology vs other aspects of impaired performance. *Neuropsychopharmacology* **41**, 2411–2420 (2016).
130. Moghaddam, B. & Krystal, J.H. Capturing the angel in “angel dust”: twenty years of translational neuroscience studies of NMDA receptor antagonists in animals and humans. *Schizophr. Bull.* **38**, 942–949 (2012).
131. Javitt, D.C., Zukin, S.R., Heresco-Levy, U. & Umbricht, D. Has an angel shown the way? Etiological and therapeutic implications of the PCP/NMDA model of schizophrenia. *Schizophr. Bull.* **38**, 958–966 (2012).
132. Wang, M. *et al.* NMDA receptors subserve persistent neuronal firing during working memory in dorsolateral prefrontal cortex. *Neuron* **77**, 736–749 (2013).
133. Skoblenick, K.J., Womelsdorf, T. & Everling, S. Ketamine alters outcome-related local field potentials in monkey prefrontal cortex. *Cereb. Cortex* **26**, 2743–2752 (2016).
134. Ma, L., Skoblenick, K., Seamans, J.K. & Everling, S. Ketamine-induced changes in the signal and noise of rule representation in working memory by lateral prefrontal neurons. *J. Neurosci.* **35**, 11612–11622 (2015).
135. Skoblenick, K. & Everling, S. NMDA antagonist ketamine reduces task selectivity in macaque dorsolateral prefrontal neurons and impairs performance of randomly interleaved prosaccades and antisaccades. *J. Neurosci.* **32**, 12018–12027 (2012).
136. Evans, S. *et al.* Performance on a probabilistic inference task in healthy subjects receiving ketamine compared with patients with schizophrenia. *J. Psychopharmacol.* **26**, 1211–1217 (2012).
137. Jones, J.A., Sponheim, S.R. & MacDonald, A.W. III. The dot pattern expectancy task: reliability and replication of deficits in schizophrenia. *Psychol. Assess.* **22**, 131–141 (2010).
138. MacDonald, A.W. III. Building a clinically relevant cognitive task: case study of the AX paradigm. *Schizophr. Bull.* **34**, 619–628 (2008).
139. Barch, D.M., Carter, C.S., MacDonald, A.W. III, Braver, T.S. & Cohen, J.D. Context-processing deficits in schizophrenia: diagnostic specificity, 4-week course, and relationships to clinical symptoms. *J. Abnorm. Psychol.* **112**, 132–143 (2003).
140. Blackman, R.K., MacDonald, A.W. III & Chafee, M.V. Effects of ketamine on context-processing performance in monkeys: a new animal model of cognitive deficits in schizophrenia. *Neuropsychopharmacology* **38**, 2090–2100 (2013).
141. Dias, E.C. *et al.* Changing plans: neural correlates of executive control in monkey and human frontal cortex. *Exp. Brain Res.* **174**, 279–291 (2006).
142. Karayiorgou, M., Simon, T.J. & Gogos, J.A. 22q11.2 microdeletions: linking DNA structural variation to brain dysfunction and schizophrenia. *Nat. Rev. Neurosci.* **11**, 402–416 (2010).
143. Stark, K.L. *et al.* Altered brain microRNA biogenesis contributes to phenotypic deficits in a 22q11-deletion mouse model. *Nat. Genet.* **40**, 751–760 (2008).
144. Sigurdsson, T., Stark, K.L., Karayiorgou, M., Gogos, J.A. & Gordon, J.A. Impaired hippocampal-prefrontal synchrony in a genetic mouse model of schizophrenia. *Nature* **464**, 763–767 (2010).
145. Fénelon, K. *et al.* Deficiency of *Dgcr8*, a gene disrupted by the 22q11.2 microdeletion, results in altered short-term plasticity in the prefrontal cortex. *Proc. Natl. Acad. Sci. USA* **108**, 4447–4452 (2011).